Hypoxic Pulmonary Vasoconstriction and Chronic Lung Disease

Erik R. Swenson, MD

Departments of Medicine and Physiology and Biophysics University of Washington VA Puget Sound Health Care System Seattle, WA Hypoxic vasoconstriction in the lung is a unique and fundamental characteristic of the pulmonary circulation. It functions in health and disease states to better preserve ventilation-perfusion matching by diverting blood flow to better ventilated regions when local ventilation is compromised. As more areas of lung become hypoxic either with high altitude or global lung disease, then hypoxic pulmonary vasoconstriction (HPV) becomes less effective in ventilation-perfusion matching and can lead to pulmonary hypertension. HPV is intrinsic to the vascular smooth muscle and its mechanisms remain poorly understood. In addition, the pulmonary vascular endothelium, red cells, lung innervation, and numerous circulating vasoactive agents also affect the strength of HPV. This review will discuss the pathophysiology of HPV and address its role in pulmonary hypertension associated with World Health Organization Group 3 diseases. When sustained beyond many hours, HPV may initiate pulmonary vascular remodeling and lead to more fixed and less oxygen-responsive pulmonary hypertension if the hypoxic stimulus is maintained.

Hypoxic pulmonary vasoconstriction (HPV) is a fundamental attribute of the pulmonary circulation, which has fascinated cardiopulmonary physiologists and clinicians since its definitive description in the cat in 1946¹ and in humans 1 year later.² It was immediately appreciated that this response to local alveolar hypoxia and hypercapnia, either alone or in combination generally occurring as result of regional hypoventilation, acts to redirect pulmonary blood flow to areas of better ventilation with their higher alveolar PO₂ and lower PCO₂. In this fashion, HPV and hypercapnic pulmonary vasoconstriction (HCPV) are potent mechanisms to better match regional perfusion (Q) to alveolar ventilation (VA) and so enhance gas exchange efficiency. If an area of regional hypoventilation is small in relation to the total pulmonary vascular bed, there is little to no increase in pulmonary artery (PA) pressure.³ When there is more global alveolar hypoxia, such as at high altitude or more extensive hypoxia with or without hypercapnia in diffuse parenchymal and airways disease, HPV still operates to optimize V_A/Q matching. However, with more of the vasculature undergoing constriction it is less effective

in this function and results in increased pulmonary vascular resistance (PVR) and pulmonary hypertension (PH).³

The presence and contribution of HPV to V_A/Q matching and PH in chronic lung diseases (World Health Organization [WHO] Group 3) and the extent to which it might be modified as part of treatment in this setting is not easily assessed. This is due to the fact that other changes in the vasculature in these conditions also increase vascular resistance. Depending on the disease and its duration and severity, these include physical destruction and loss of vascular bed with a decrease in total perfused cross-sectional area, hyperinflation such as with emphysema, reduction in local tonic vasodilator generation and/or increase in vasoconstrictor mediator production, and remodeling of existing vessels with increased smooth muscle mass and perivascular thickening leading to luminal narrowing.

In this review, the present understanding of HPV in the normal and diseased lung will be discussed with the goal of understanding its contribution to WHO Group 3 PH and its potential to be targeted therapeutically or be altered by treatments for these conditions.

Key Words—arterial hypoxemia, high altitude, hypercapnia, hypoxia, normoxia Correspondence: eswenson@u.washington.edu

CHARACTERIZATION OF HPV

Increases in PVR and PA pressure on ascent to high altitude or exposure to normobaric hypoxia universally occur in humans and other mammals. HPV can be detected with elevations in altitude as low as 1600-2500 m or with reductions in F_IO₂ to 0.15-0.18.^{4,5} The magnitude of HPV (Figure 1) can vary almost 5-fold among healthy individuals,⁶ and among species (Figure 2) in part related to total pulmonary vascular smooth $muscle^{6,\overline{7}}$ and with duration of hypoxia (Figure 3) from minutes to several days.⁷⁻⁹ HPV is the earliest mechanism that elevates PA pressure and PVR with hypoxic or high-altitude exposure. Ultimately, other mechanisms (perhaps partly in reaction to the first elevation of pressure initiated by HPV along with greater cardiac output) such as activation of pressure-independent hypoxiasensitive inflammatory and proliferative pathways¹⁰ may contribute to sustained PVR elevation and vascular remodeling. The process of remodeling is initiated as early as several hours at the level of new gene transcription, such as for collagen and other growth factors,¹¹ and is generally established within days to weeks of continuous alveolar hypoxia.12-14

Disclosures: Dr Swenson reports financial relationships with Cardeas, Boehringer Ingelheim, and Novartis.

The ability to reverse the acute effects of HPV by restoration of normoxia progressively diminishes with sustained



Figure 1. HPV variability as assessed by PA systolic pressure response in normal humans to 4 hours of moderate hypoxia. Subjects noted by solid lines are subjects susceptible to HAPE and show exaggerated HPV, while subjects without HAPE susceptibility (interrupted lines) have lower HPV. (Grunig et al. *J Am Coll Cardiol*, 2000.)

hypoxic exposure. This decline in reversibility has been demonstrated as early as 8 hours⁹ and progressing through 1 to 3 days,^{15,16} (Figure 4) and becomes more pronounced after 1 to several weeks of hypoxic exposure.^{8,17,18} Although changes in inspired oxygen remain widely used to assess HPV, changes in arterial oxygenation and acid-base status always follow an alteration in inspired oxygen, so that systemic effects such as changes in central nervous system (CNS) and autonomic nervous activity might also contribute to the final pulmonary vascular response. It would be useful to employ a truly selective HPV inhibitor or stimulator in vivo rather than use changes in inspired oxygen, but all

available pulmonary vasoactive agents have actions elsewhere in the circulation and brain, making them less than ideal for this purpose.

Lowland species with stronger acute HPV tend to develop greater PH with chronic hypoxia than animals with weaker HPV.¹² Whether humans with stronger HPV develop greater PH with chronic hypoxia or other conditions predisposing to PH has never been studied, but such a characteristic might underlie those often labeled as having "out-ofproportion" hypertension in the face of subsequent development of obstructive sleep apnea, heart failure, emphysema, and fibrotic lung disease. These conditions are more prevalent in older patients, and the impact of aging on HPV is also unknown.

The critical PO_2 at the level of the pulmonary arteriolar smooth muscle which initiates HPV in a lung region or the whole lung is a summation of the effects of alveolar PO₂ as set by inspired PO₂ and the ventilation-perfusion (V_A/Q) ratio, the bronchial arterial PO₂, and the mixed venous PO2.3 Because the bronchial arterial circulation perfuses the vaso vasorum of the pulmonary arteries and arterioles, systemic arterial PO2 will also influence HPV. Separate perfusion of the bronchial artery in the sheep with deoxygenated blood, while alveolar PO₂ and systemic PO₂ were held constant, led to an increase in PA pressure.¹⁹ In animal studies in which it is possible to control and hold systemic arterial and alveolar PO2 constant, reductions in mixed venous PO2 sensed in the PA cause vasoconstriction.^{20,21} The importance of mixed venous PO2 as a factor in HPV may be magnified with exercise, when mixed venous PO_2 falls to very low tensions as a result of high tissue oxygen extraction and greater arterial hypoxemia than at rest, but of the 3 contributions mixed venous PO₂ likely has the least influence.

MECHANISMS OF ACUTE HPV

HPV is a complex process with elements of its expression arising from multiple points in the neuro-cardiopulmonary axis, with variation in intensity and mechanisms over time.²² In addition to the intrinsic hypoxic response of the pulmonary vasculature that can be elicited in isolated pulmonary vascular smooth muscle cells and vessels, there are numerous extrinsic modulating influences sensitive to oxygen in vivo that include the vascular endothelial cells, red cells, chemoreceptors, autonomic nervous system, and lung innervation. The pulmonary circulation response to hypoxia is characterized by contraction of smooth muscle cells of the small pulmonary arterioles and veins of diameter less than 900 μ m; the veins account for approximately 20% of the total increase in PVR.^{23,24} At a regional level within the lung vasculature the magnitude of HPV may not be equivalent in all areas or static over time.²⁵⁻²⁸ As a consequence of this

unevenness of regional HPV, some areas of the vasculature may be more perfused than others if they have a lower HPV response. This appears to be the case in those with a stronger global HPV response and susceptibility to high altitude pulmonary edema (HAPE).26,27 Although it is not generally thought that hypoxia acts at the microvascular or acinar level, pulmonary capillary endothelial cells respond to hypoxia with membrane depolarization,²⁹ and this signal is propagated upstream and possibly downstream to resistance arterioles and venules. As vet, no evidence has been found for capillary constriction with hypoxia,³⁰ despite evidence that other vasoconstrictors are active at this level and in surrounding parenchymal perivascular cells that contain actin and myosin microfilaments.31

HPV in intact animals and humans appears to be fully expressed within 6 to 8 hours and has several temporal components. The first occurs within 5 minutes with a half-time of about 30-90 seconds.³²⁻³⁵ A second phase of greater pressure elevation (almost double) is evident in humans and plateaus at 2 hours.³⁵ In animal studies, further elevation of pressure develops over the next 6 to 8 hours.³² This has been confirmed in studies of isolated pulmonary arteries, lungs, or vascular smooth muscle cells showing a third phase taking upward of 8 hours.³⁶ The mechanisms behind these differing time phases and differences between in vivo and isolated lung and vessel investigations have not been well studied, but the isolated vessel studies suggest the first phase is intrinsic calciumdependent smooth muscle contraction, with the later phases representing the summation of numerous other modulating influences acting on the smooth muscle^{22,36} in a calcium concentrationindependent fashion as discussed below. All of these differing hypoxic responses are fully and immediately reversible with return to normoxia if hypoxia is not extended beyond several hours.

HPV at the Level of the Vascular Smooth Muscle

There are several mechanisms involved in HPV that are activated in parallel or sequentially, leading to a critical increase



Figure 2. HPV in a variety of mammals showing baseline differences in normoxic mean PA pressure and increases with acute hypoxia. (Reeves et al. *Int Rev Physiol*, 1979.)

of intracellular calcium and/or an enhanced calcium sensitivity of the actinmyosin that initiates contraction, 13,22 a response opposite to that which occurs in the systemic vasculature. Intracellular calcium concentration is increased by hypoxia-mediated inhibition of several potassium channels, leading to membrane depolarization and extracellular calcium entry through L-type channels, and a release of calcium from the sarcoplasmic reticulum (SR), with further influx through store-operated calcium channels (SOCC), receptor-operated calcium channels (ROCC), and transient receptor potential channel 6 (TRPC6). Figure 5 depicts the very complicated multiple pathways by which intracellular calcium in pulmonary vascular smooth muscle is quickly altered by hypoxia to initiate HPV. In addition, sensitivity to calcium of the contractile elements is enhanced via a hypoxia-induced increase

in Rho-kinase activity.³⁷ The change in oxygen tension that stimulates these components of HPV is signaled by an alteration in the redox status of the smooth muscle cells.^{13,38} Whether an increase or a decrease of reactive oxygen species (ROS) is responsible for HPV signal transduction is still under debate, but a stronger case is emerging that hypoxia increases mitochondrial ROS generation as an upstream signal for HPV.³⁸ It is clear that high-altitude exposure increases stable circulating markers of ROS production, and persons with higher HPV appear to generate more ROS and less bioactive vasodilating nitric oxide (NO) species across the lung.³⁹

Endothelium-Dependent Modulation of HPV

The pulmonary vascular endothelium generates a variety of vasoactive medi-



Figure 3. Species variability in severity of PH during exposure to chronic hypoxia. (Reeves et al. *Int Rev Physiol*, 1979.)

ators that act in a paracrine fashion on the surrounding vascular smooth muscle cells. These include NO and prostacyclin as vasodilators, and endothelin-1 acting as a vasoconstrictor via binding to endothelin-A receptors and a vasodilator by binding to endothelin-B receptors causing NO release.³⁶ Isolated human PA endothelial cells exposed to 3% oxygen produce more hydrogen peroxide and thus may also be a source for ROS that initiate HPV.⁴⁰ The endothelium also produces carbon monoxide (CO) via heme-oxygenase-2,41 which is upregulated by hypoxia.^{42,43} CO dilates vessels by activating guanylate cyclase to generate cyclic guanosine monophosphate (GMP) in a manner similar to NO. Hydrogen sulfide (H_2S) , a strong reducing agent, generated in hypoxia is vasoconstricting in the pulmonary circulation by several not yet fully quantified

mechanisms.⁴⁴ It should be noted that many of these "gaso-transmitters" alter the concentrations of each other, making it difficult to assess the contribution of each to HPV modulation.^{45,46}

Erythrocyte-Dependent Modulation of HPV

Red cells may contribute to HPV and pulmonary pressures in several ways. Although hypoxia-mediated decrease in deformability might reduce flow and increase measured vascular resistance,^{47,48} direct measurements of human and other mammalian red cells over a range of PO_2 from 120 to 47 mm Hg show no evidence of significant deformability changes.⁴⁹ With elevations in hematocrit with altitude, pulmonary vascular pressures increase. This is partly due to increased blood viscosity and direct increases in lung vascular resistance as

shown by hemodilution studies at high altitude in patients with chronic mountain sickness⁵⁰ and in animal studies.⁵¹ Red cell-mediated changes in PVR with hypoxia represent a balance between those effects that are vasodilating and others that are vasoconstricting. Direct endothelial cell NO scavenging by oxyhemoglobin⁵² and ROS generation by hypoxic red cells⁵³ will enhance HPV. In contrast, the oxygenation dependent behavior of red cells and hemoglobin that lead to s-nitrosothiol release⁵⁴ and NO generation from nitrite with hemoglobin desaturation" will blunt HPV. Additionally, deoxygenated red cells also release adenosine triphosphate (ATP), which activates endothelial cell NO production via purinergic receptor binding⁵⁶ and so act in a vasodilating fashion. Finally, recent evidence that red cells themselves express the endothelial isozyme of nitric oxide synthase (eNOS) and are able generate NO that escapes intracellular hemoglobin binding⁵⁷ needs to be considered. Similar to the various and sometimes competing interactions of endothelial cell vasoactive mediators on HPV, the contribution of red cells is similarly complicated and the net result on PVR may vary depending on the degree and duration of hypoxia.

Neurohumoral-Dependent Modulation of HPV

The lung vasculature is innervated by sympathetic noradrenergic fibers from the large conduit arteries and veins down to 50 μ m vessels in larger species such as man and dogs, but much less so in smaller species.⁵⁸ In addition to release of norepinephrine with sympathetic activation causing vasoconstriction via alpha-1 adrenergic receptors on vascular smooth muscle, there is release of other opposing vasodilating neurotransmitters such as neuropeptide Y and vasoactive intestinal peptide.58 Additionally, there is opposing NO-dependent vasodilating parasympathetic innervation.⁵⁹ Arterial PO₂ is gauged by the peripheral chemoreceptors, which project afferents to the medullary cardiovascular control areas in the brain stem in addition to the respiratory control center, activating both

parasympathetic and sympathetic outflow to the lung. Denervation of the carotid bodies and loss of afferent input from the peripheral chemoreceptors increases HPV.^{60,61} The efferent arc of this response is not well defined but is conveyed by the vagus nerve. Vagotomy reduces HPV.^{62,63} Studies using atropine and propranolol suggest that vasodilating parasympathetic activity is more dominant than sympathetic activity in HPV inhibition.^{63,64} Other data suggest a stronger sympathetic contribution.⁶⁵

In regard to neurohumoral mediation of HPV, susceptibility to HAPE is characterized by a very exaggerated HPV⁶⁶ and a much greater generalized sympathetic nervous system activation to hypoxia.^{67,68} However, not all studies find evidence for neural modulation of HPV.⁶⁹ The reason for this discrepancy is not clear, but those studies finding no effect on HPV have employed receptor blocking drugs rather than neural pathway interruption. It is entirely possible that peripheral chemoreceptormediated modulation of HPV may involve other neurotransmitter release via the lung innervation besides catecholaminergic or cholinergic agonists as described above. In humans, the association of stronger hypoxic ventilatory response (HVR), which is almost wholly a peripheral chemoreceptor mediated response, with weaker HPV supports the majority of the animal work.70

The pulmonary vasculature expresses adrenergic and cholinergic receptors, as well as other receptors, including those for thyroxine, angiotensin II, adenosine, natriuretic peptides, and estrogen. Thus it can respond to circulating vasoactive mediators with dilation by epinephrine via beta-2 receptors,⁵⁸ estrogens⁷¹ and natriuretic peptides,⁷² and constriction with angiotensin,⁷³ adenosine,⁷⁴ and thyroxine.⁷⁵ The full neurohumoral component of the lung vascular response to hypoxia is often neglected in discussions of HPV.

Other Modulating Influences on HPV

Individual genetic background⁷⁶⁻⁷⁸ and a history of familial susceptibility to HAPE or PH⁷⁹⁻⁸¹ also contribute to the strength of HPV. Acid-base status and carbon dioxide have a considerable



Figure 4. Time course in PA pressure (systolic, mean, and diastolic) and wedge pressure in humans with acute hypoxia showing lack of complete resolution of HPV with return to normoxia after 8 hours. (Dorrington et al. *J Appl Physiol*, 1997.)

influence on HPV, with alkalosis and hypocapnia both diminishing HPV and hypercapnia increasing HPV. 82,83 Thus subjects with stronger ventilatory responses to hypoxia will not only maintain higher alveolar PO₂, but also will have less HPV due to their greater hypocapnic alkalemia at any given altitude or F₁O₂. Increasing lung volume by positive end-expiratory pressure in the range of 8-10 cmH₂O does not reduce HPV.⁸⁴ Pre-existing high arterial wall tension also diminishes HPV.85 Lastly, animal studies with low-grade infection or inflammation show that circulating and locally produced inflammatory leukotrienes, thromboxanes and cytokines, (ie, tumor necrosis factor, interleukin-6),⁸⁶⁻⁸⁹ or activation of their receptors in

the vasculature⁹⁰ appear to modulate HPV (both negatively and positively).

Hypoxia-Regulated Gene Transcription Factors and HPV

The study of HPV continues to identify new sensing, signaling, and effector mechanisms and pathways. The most recent are the hypoxia-inducible factors (HIFs), transcription factors that alter the gene expression over 1000 genes involved in promoting tolerance to hypoxia.⁹¹ Additionally, HIF activates a number of inflammatory signaling molecules such as nuclear factor kappa beta.¹⁰ In this fashion, hypoxia and inflammation may be inextricably linked in chronic lung diseases. In 2 rat strains with differing pulmonary hypoxic



Figure 5. Diagram of pulmonary arterial smooth muscle cell intracellular calcium ([Ca²⁺]i) pathways with acute hypoxia. Pathways that increase ([Ca²⁺]i are shown on the left, while those that decrease ([Ca²⁺]i are on the right. Hypoxia can activate (green) or inhibit (red) these pathways. Whether these effects are probable, possible, or speculative is indicated by solid, dashed, and dotted lines, respectively, as shown in the key at the bottom. With respect to plasma membrane and associated cytosolic signals, TASK-1 is TWIK-related acid-sensitive channel-1;VOCC, K_V, Cl_{Ca}, SOCC, NSCC, and ROCC indicate voltage-operated Ca²⁺, voltagedependent K⁺, calcium-dependent Cl, store-operated Ca²⁺, nonselective cation, and receptor-operated Ca²⁺ channels, respectively. NCX, Na-Ca exchanger; A, agonist; R, receptor; PLC, phospholipase C; PIP₂, phosphatidylinositol 4,5-bisphosphate; IP₃, inositol 1,4,5-trisphosphate; DAG, diacylglycerol; PKC, protein kinase C; PMCA, plasma membrane Ca2_-ATPase. With respect to sarcoplasmic reticulum (SR) and associated cytosolic signals, SERCA is sarcoplasmic-endoplasmic reticulum ATPase, IP3R is IP3 receptor, RyR is ryanodine receptor, STIM1 is stromal interaction molecule 1, and cADPR is cyclic ADP ribose. With respect to lysosome-like organelles (LLO) and associated cytosolic signals, NAADP is nicotinic acid adenine dinucleotide phosphate, HCX is H-Ca exchanger, and HA is H⁺-ATPase. Mito, mitochondria. (Sylvester et al. Physiol Rev, 2012.)

responses, HIF-1 activity and HIFmediated protein expression were higher in the strain with greater PH.⁹² In contrast, mice with heterozygous HIF-1 alpha deficiency have weaker acute and chronic hypoxic responses in isolated pulmonary vascular smooth myocytes and pulmonary vessels than wild-type mice.^{93,94} Further supporting pharmacological evidence for HIF-1 alpha mediation of HPV was demonstrated in mice by reduction in hypoxic PH⁹⁵ with digoxin, a known inhibitor of HIF-1 alpha transcriptional activity.⁹⁶ At present it is not fully clear how HIFdependent gene transcription affects HPV, but it likely involves upregulation of TRPC on the vascular smooth muscle cell membrane⁹⁷ and alterations in pulmonary vascular smooth muscle calcium signaling.⁹⁵ Iron is emerging as a critical element in HPV and pulmonary vascular changes with hypoxia. Iron supplementation and iron chelation reduce and increase HPV respectively,^{98,99} possibly via altered HIF metabolism¹⁰⁰ involving prolyl hydroxylases, the O₂-sensitive enzymes that degrade HIF and require iron. HIFmediated gene transcription also drives much of the longer-term remodeling of the vasculature.¹⁴

RELEVANCE OF HPV IN HEALTH

At low altitudes where humans evolved, it would appear that the sensitivity to oxygen of the lung vasculature evolved along with HCPV as mechanisms¹⁰¹ to shift blood flow from poorly or nonventilated lung regions with localized airway or airspace pathology in post-fetal life to better ventilated and healthy areas as elegantly advanced by von Euler and Liljestrand¹ in their landmark paper. Based on whole lung pulmonary vascular responses to changes in alveolar PO2 and PCO₂, Dorrington et al¹⁰¹ modelled that improvements in V_A/Q matching and gas exchange by HPV are most important in the lower range of V_A/Q_{-} (0.01 to 1.0), and that HCPV has its greatest impact in the V_A/Q ratio of 1 to 100. The ability of both HPV and HCPV to divert blood flow and minimally raise PA pressure are more effective when the area of VA/Q mismatching is smaller.³ From an evolutionary perspective, HPV may have conferred a survival (and ultimately a reproductive) advantage for individuals with severe pneumonia or thoracic trauma with acute pneumothorax by limiting the degree of severe life threatening shunt-induced hypoxemia. This may still be the case even in the modern clinical era of effective antibiotics and surgery before patients can be treated.¹⁰²

Alternatively, others have argued it may be simply a vestige of fetal existence. In this regard, HPV maintains a high vascular resistance to limit blood flow in the nonventilated lung (in combination with a patent ductus arteriosis and foramen ovale) to allow an 80% to 90% right to left shunt to provide more blood flow to the placenta and better oxygenated blood to the developing brain.¹⁰³ However, many other aspects of the fetal lung also contribute to higher PVR, including its liquid-filled nonventilated high volume

Downloaded from https://prime-pdf-watermark.prime-prod.pubfactory.com/ at 2025-06-24 via free access

state, lack of surfactant, relative hypercapnia and acidosis, a limited slower growing vascular bed relative to the faster growing airway and parenchymal structure, lesser endothelial vasodilator generation, greater endothelial vasoconstrictor production, and lack of bronchial epithelial NO generation.^{103,104} In fact, HPV in the fetal lung does not appear until the middle of the third trimester of gestation. Thus, it would appear to reduce PVR and prepare the pulmonary circulation to accommodating the entire cardiac output at birth as the ductus arteriosis closes and the lungs are ventilated and assume gas exchange duties from the placenta.¹⁰⁴ In this sense, HPV should perhaps more correctly be renamed "oxygen-dependent vasodilation." If strong HPV is an evolutionary advantage in utero, then one might predict a fetal survival disadvantage in Tibetans, who have much lower HPV as adults than other populations.¹⁰⁵ Yet, birth rates and neonatal survival in this population exceed those of newcomers to high altitude.¹⁰⁶

RELEVANCE OF HPV IN CHRONIC LUNG DISEASE

In the setting of chronic lung diseases, several questions regarding HPV are relevant. The first is whether it is present and what is its magnitude. The second is how useful is HPV in maintaining as optimal state of gas exchange as possible. The third is what benefit or harm is realized with therapies that either directly alter HPV or alter it as a consequence of targeting some other aspect of the disease.

In answering the first and second questions, if the model of chronic global hypoxia^{107,108} such as that occurring with long-term high-altitude exposure is any answer, then the finding that after several weeks at high altitude there is little pulmonary vasodilation with breathing oxygen would suggest HPV should not be present to any great extent in chronic hypoxic lung diseases.^{8,105} Some patients with chronic obstructive pulmonary disease (COPD) and chronic bronchitis given high levels of inspired oxygen acutely show deterioration in V_A/Q matching suggestive of inhibition of HPV,¹⁰⁹ but this has not been shown in every case.¹¹⁰ Although these data

and data from other studies have been used to support the idea that HPV is contributing to the high vascular tone, in studies with right heart catheterization there is minimal reduction in PH with supplemental oxygen therapy either acutely or chronically in most patients.¹¹² This apparent paradox might be explained either by there being only small regions of lung having any HPV, such that gas exchange deterioration still takes place, but reduction in overall PA pressure is minimal. A second possibility is that simultaneous increase in local carbon dioxide brought about by an increase in blood flow with release of HPV¹¹³ in these areas leads to counteracting HCPV and limits the fall of pulmonary artery pressure. Despite the equivocal salutary effects of short-term oxygen, it is clearly established that chronic supplemental oxygen extends life in hypoxemic COPD patients and that this is associated with a mild improvement in pulmonary hemodynamics in those using continuous oxygen.¹¹⁴ In patients exhibiting a significant drop in mean PA pressure of >5 mm Hg the benefits were greatest.¹¹⁵ The benefits of oxygen therapy are multiple and stem largely from improvements in systemic oxygenation. However, the pulmonary vascular effects of oxygen in the long run may be related to HPV in much the same way that all models of chronic hypoxic PH in animals and in humans relocating from high altitude to sea level ultimately show regression of PH after return to normoxia.^{107,108}

In interstitial lung disease the story is different. Two studies have shown no significant vasodilator response or change in V_A/Q matching with 100% oxygen,^{116,117} and chronic home oxygen administration does not alter mortality in fibrotic lung diseases.¹¹⁸ Therefore, from these data it appears that HPV does not contribute greatly to PH in interstitial lung disease.¹¹⁹

HPV can be decreased for treatment purposes by a variety of pharmacological agents that act on many of the endothelial cell-derived modulators of PVR, signal transduction pathways, and gene transcription discussed above, including NO, nitrates, calcium channel blockers, phosphodiesterase 5 inhibitors, endothelin receptor blockers, prostacyclin analogs, soluble guanylate cyclase (sGC) activators, angiotensin converting enzyme inhibitors, and some carbonic anhydrase inhibitors, such as acetazolamide.²² While these drugs certainly inhibit HPV at high altitude and some are quite useful to prevent and treat HAPE and high-altitude PH,⁶⁶ it must be appreciated that none of these agents are truly specific HPV inhibitors, except perhaps for acetazolamide.¹²⁰ Their pressure-lowering effects act on intracellular calcium signaling, mediator release, or receptor engagement, some of which may be common to HPV.

Several of these drugs that have been tested in patients with COPD and idiopathic pulmonary fibrosis (IPF) (as will be discussed in the 2 accompanying articles in this issue) may impair gas exchange efficiency by inhibiting HPV and/or by general vasodilation more in areas of shunt or low V_A/Q . For instance, with oral sildenafil in COPD, PA pressure is lowered at equivalent exercise intensity,¹²¹ but in some arterial PO_2 falls. In those that derive an exercise and pressure-lowering effect, the drop in oxygenation could be likely prevented by small increases in their supplemental oxygen flow rate. Whether this is a tenable approach and might increase exercise capacity requires formal testing. (Note added in proof: A recent study by Blanco et al. (Eur Respir J. 2013;42:982-99) showed no benefit of sildenafil to a comprehensive pulmonary rehabilitation program in exercise endurance or quality of life.)

Lastly, the adverse effect of giving these agents orally might be mitigated by giving them by inhalation in order to vasodilate preferentially in the better ventilated regions and not worsen V_A/Q mismatch such as with iloprost.^{122,123}

CONCLUSION

The search for more potent and selective vasodilators for the treatment of nonhypoxic forms of PH grows apace, and it is likely that most will have the ability to inhibit HPV. It may be useful in selected patients without an obvious ventilatory limitation at maximal exercise to measure how much both oxygen and medications lower PA pressure and restrict their use to those with reductions in PVR associated with increased functional capacity or decreased dyspnea while adding or increasing supplemental oxygen as needed to maintain acceptable arterial oxygenation levels.

There is considerable diversity among WHO Group 3 PH patients and within the individual diagnostic subsets comprising this group. While HPV may play a variable role in the pathogenesis of PH in Group 3 patients, and treatment of hypoxia remains an important therapeutic consideration, the heterogeneity of this population poses significant challenges for development of effective treatment.

Multiple pathways associated with HPV, HCPV, other V_A/Q matching mechanisms, hyperinflation, inflammation, vascular remodeling, and parenchymal loss contribute to the development of PH and pose significant challenges for identification and evaluation of potential therapeutic agents. Understanding these mechanisms and identifying patient groups where similar pathways predominate is a critical component in the evolution of treatment for WHO Group 3 PH.

References

1. von Euler US, Liljestrand G. Observations on the pulmonary arterial blood pressure in the cat. *Acta Physiol Scand.* 1946;12:301-320.

2. Motley HL, Cournand A, Werko L, et al. The influence of short periods of induced acute anoxia upon pulmonary artery pressures in man. *Am J Physiol.* 1947;150(2):315-320.

3. Marshall BE, Marshall C. A model for hypoxic constriction of the pulmonary circulation. *J Appl Physiol (1985).* 1988;64(1):68-77.

4. Levine BD, Zuckerman J, deFillipi CR. The effect of high-altitude exposure in the elderly: the Tenth Mountain Division study. *Circulation* 1997; 96(4):1224-1232.

5. Smith TG, Talbot NP, Chang RW, et al. Pulmonary artery pressure increases during commercial air travel in healthy passengers. *Aviat Space Environ Med.* 2012;83(7):673-676.

6. Tucker A, McMurtry IF, Reeves JT, Alexander AF, Will DH, Grover RF. Lung vascular smooth muscle as a determinant of pulmonary hypertension at high altitude. *Am J Physiol.* 1975; 228(3):762-767.

7. Grünig E, Mereles D, Hildebrandt W, et al. Stress Doppler echocardiography for identification of susceptibility to high altitude pulmonary edema. *J Am Coll Cardiol.* 2000;35(4):980-987. 8. Groves BM, Reeves JT, Sutton JR, et al. Operation Everest II: elevated high-altitude pulmonary resistance unresponsive to oxygen. *J Appl Physiol (1985).* 1987;63(2):521-530.

9. Dorrington KL, Clar C, Young JD, Jonas M, Tansley JG, Robbins PA. Time course of the human pulmonary vascular response to 8 hours of isocapnic hypoxia. *Am J Physiol.* 1997;273(3 Pt 2):H1126-H1134.

10. Voelkel NF, Mizuno S, Bogaard HJ. The role of hypoxia in pulmonary vascular diseases: a perspective. *Am J Physiol Lung Cell Mol Physiol.* 2013;304(7):L457-L465.

11. Berg JT, Breen EC, Fu Z, Mathieu-Costello O, West JB. Alveolar hypoxia increases gene expression of extracellular matrix proteins and platelet-derived growth factor-B in lung parenchyma. *Am J Respir Crit Care Med.* 1998;158(6): 1920-1928.

12. Grover RF. The fascination of the hypoxic lung. *Anesthesiol.* 1985;63(6):580-582.

13. Sommer N, Dietrich A, Schermuly RT, et al. Regulation of hypoxic pulmonary vasoconstriction: basic mechanisms. *Eur Respir J.* 2008;32(6):1639-1651.

14. Welsh DJ, Peacock AJ. Cellular responses to hypoxia in the pulmonary circulation. *High Alt Med Biol.* 2013;14(2):111-116.

15. Kronenberg RS, Safar P, Leej, et al. Pulmonary artery pressure and alveolar gas exchange in man during acclimatization to 12,470 ft. *J Clin Invest.* 1971;50(4):827-837.

16. Maggiorini M, Melot C, Pierre S, et al. High-altitude pulmonary edema is initially caused by an increase in capillary pressure. *Circulation*. 2001;103(16):2078-2083.

17. Canepa A, Chavez R, Hurtado A, Rotta A, Velasquez T. Pulmonary circulation at sea level and at high altitudes. *J Appl Physiol.* 1956;9(3): 328-335.

18. Dubowitz G, Peacock AJ. Pulmonary artery pressure in healthy subjects at 4250 m measured by Doppler echocardiography. *Wilderness Environ Med.* 2007;18(4):305-311.

 Marshall BE, Marshall C, Magno M, Lilagan P, Pietra GG. Influence of bronchial arterial PO₂ on pulmonary vascular resistance. *J Appl Physiol (1985)*. 1991;70(1):405-415.

20. Hyman AL, Higashida RT, Spannhake EW, Kadowitz PJ. Pulmonary vasoconstrictor responses to graded decreases in precapillary blood PO₂ in intact-chest cat. *J Appl Physiol Respir Environ Exerc Physiol.* 1981;51(4):1009-1016.

 Pease RD, Benumof JL, Trousdale FR. PAO2 and PVO2 interaction on hypoxic pulmonary vasoconstriction. *J Appl Physiol Respir Environ Exerc Physiol.* 1982;53(1):134-139.
 Sylvester JT, Shimoda LA, Aaronson PI,

Ward JP. Hypoxic pulmonary vasoconstriction. *Physiol Rev.* 2012;92(1):367-520.

23. Hakim TS, Michel RP, Chang HK. Site of pulmonary hypoxic vasoconstriction studied with arterial and venous occlusion. *J Appl Physiol Respir Environ Exerc Physiol.* 1983;54(5):1298-1302.

24. Audi SH, Dawson CA, Rickaby DA, Linehan JH. 1991. Localization of the sites of pulmonary vasomotion by use of arterial and venous occlusion. J Appl Physiol (1985). 1991;70(5):2126-2136.

25. Dawson A. Regional pulmonary blood flow in sitting and supine man during and after acute hypoxia. *J Clin Invest*. 1969;48(2):301-310.

 Hopkins SR, Garg J, Bolar DS, Balouch J, Levin DL. Pulmonary blood flow heterogeneity during hypoxia and high-altitude pulmonary edema. *Am J Respir Crit Care Med.* 2005;171(1): 83-87.

27. Dehnert C, Risse F, Ley S, et al. Magnetic resonance imaging of uneven pulmonary perfusion in hypoxia in humans. *Am J Respir Crit Care Med.* 2006;174(10):1132-1138.

28. Asadi AK, Cronin MV, Sá RC, et al. Spatial-temporal dynamics of pulmonary blood flow in the healthy human lung in response to altered F_1O_2 . *J Appl Physiol (1985)*. 2013;114(1): 107-118.

29. Wang L, Yin J, Nickles HT, et al. Hypoxic pulmonary vasoconstriction requires connexin 40-mediated endothelial signal conduction. *J Clin Invest*. 2012;122(11):4218-4230.

30. Conhaim RL, Burt Olson E Jr, Vidruk EH, et al. Acute hypoxia does not alter inter-alveolar perfusion distribution in unanesthetized rats. *Respir Physiol Neurobiol.* 2008;160(3):277-283.

31. Watson KE, Dovi WF, Conhaim RL. Evidence for active control of perfusion within lung microvessels. *J Appl Physiol (1985)*. 2012;112(1): 48-53.

32. Vejlstrup NG, O'Neill M, Nagyova B, Dorrington KL. Time course of hypoxic pulmonary vasoconstriction: a rabbit model of regional hypoxia. *Am J Respir Crit Care Med.* 1997;155(1): 216-221.

33. Morrell NW, Nijran KS, Biggs T, Seed WA. Magnitude and time course of acute hypoxic pulmonary vasoconstriction in man. *Respir Physiol.* 1995;100(3):271-281.

34. Deem S, Hedges RG, Kerr ME, Swenson ER. Acetazolamide reduces hypoxic pulmonary vasoconstriction in isolated perfused rabbit lungs. *Respir Physiol.* 2000;123(1-2):109-119.

35. Talbot NP, Balanos GM, Dorrington KL, Robbins PA. Two temporal components within the human pulmonary vascular response to approximately 2 h of isocapnic hypoxia. *J Appl Physiol* (1985). 2005;98(3):1125-1139.

36. Aaronson PI, Robertson TP, Ward JP. Endothelium-derived mediators and hypoxic pulmonary vasoconstriction. *Respir Physiol Neurobiol.* 2002;132(1):107-120.

Weigand L, Shimoda LA, Sylvester JT.
 2011. Enhancement of myofilament calcium sensitivity by acute hypoxia in rat distal pulmonary arteries. *Am J Physiol Lung Cell Mol Physiol.* 2011; 301(3):L380-L387.

38. Schumacker PT. Lung cell hypoxia: role of mitochondrial reactive oxygen species signaling in triggering responses. *Proc Am Thorac Soc.* 2011; 8(6):477-484.

39. Bailey DM, Dehnert C, Luks AM, et al. High-altitude pulmonary hypertension is associated with a free radical-mediated reduction in pulmonary nitric oxide bioavailability. *J Physiol.* 2010; 588(Pt 23):4837-4847.

40. Irwin DC, McCord JM, Nozik-Grayck E, et

al. A potential role for reactive oxygen species and the HIF-1alpha-VEGF pathway in hypoxiainduced pulmonary vascular leak. *Free Radic Biol Med.* 2009;47(1):55-61.

41. Zhang F, Kaide JI, Yang L, et al. CO modulates pulmonary vascular response to acute hypoxia: relation to endothelin. *Am J Physiol Heart Circ Physiol.* 2004;286(1):H137-H144.

42. Motterlini R, Foresti R, Bassi R, Calabrese V, Clark JE, Green CJ. Endothelial heme oxygenase-1 induction by hypoxia. Modulation by inducible nitric-oxide synthase and S-nitrosothiols. *J Biol Chem.* 2000;275(18):13613-13620.

43. Llanos AJ, Ebensperger G, Herrera EA, et al. The heme oxygenase-carbon monoxide system in the regulation of cardiorespiratory function at high altitude. *Respir Physiol Neurobiol.* 2012;184(2): 186-191.

44. Madden JA, Ahlf SB, Dantuma MW, Olson KR, Roerig DL. Precursors and inhibitors of hydrogen sulfide synthesis affect acute hypoxic pulmonary vasoconstriction in the intact lung. *J Appl Physiol (1985).* 2012;112(3):411-418.

45. Skovgaard N, Gouliaev A, Aalling M, Simonsen U. The role of endogenous H2S in cardiovascular physiology. *Curr Pharm Biotechnol.* 2011;12(9):1385-1393.

46. Evans AM, Hardie DG, Peers C, Mahmoud A. Hypoxic pulmonary vasoconstriction: mechanisms of oxygen-sensing. *Curr Opin Anaesthesiol*. 2011;24(1):13-20.

47. Palareti G, Coccheri S, Poggi M, Tricarico MG, Magelli M, Cavazzuti F. Changes in the rheologic properties of blood after a high altitude expedition. *Angiology.* 1984;35(7):451-458.

48. Hakim TS, Macek AS. Role of erythrocyte deformability in the acute hypoxic pressor response in the pulmonary vasculature. *Respir Physiol.* 1988; 72(1):95-107.

49. Kaniewski WS, Hakim TS, Freedman JC. Cellular deformability of normoxic and hypoxic mammalian red blood cells. *Biorheology*. 1994;31(1): 91-101.

50. Manier G, Guenard H, Castaing Y, Varene N, Vargas E. Pulmonary gas exchange in Andean natives with excessive polycythemia–effect of hemodilution. *J Appl Physiol (1985).* 1988;65(5): 2107-2117.

51. Kerbaul F, Van der Linden P, Pierre S, et al. Prevention of hemodilution-induced inhibition of hypoxic pulmonary vasoconstriction by N-acetylcysteine in dogs. *Anesth Analg.* 2004;99(2): 547-551.

52. Deem S, Swenson ER, Alberts MK, Hedges RG, Bishop MJ. Red-blood-cell augmentation of hypoxic pulmonary vasoconstriction: hematocrit dependence and the importance of nitric oxide. *Am J Respir Crit Care Med.* 1998;157(4 Pt 1): 1181-1186.

53. Kiefmann R, Rifkind JM, Nagababu E, Bhattacharya J. Red blood cells induce hypoxic lung inflammation. *Blood.* 2008;111(10): 5205-5214.

54. Gaston B, Singel D, Doctor A, Stamler JS. S-nitrosothiol signaling in respiratory biology. *Am J Respir Crit Care Med.* 2006;173(11):1186-1193. 55. Crawford JH, Isbell TS, Huang Z, et al. Hypoxia, red blood cells, and nitrite regulate NO-dependent hypoxic vasodilation. *Blood.* 2006; 107(2):566-574.

56. Sprague RS, Olearczyk JJ, Spence DM, Stephenson AH, Sprung RW, Lonigro AJ. Extracellular ATP signaling in the rabbit lung: erythrocytes as determinants of vascular resistance. *Am J Physiol Heart Circ Physiol.* 2003;285(2):H693-H700.

57. Cortese-Krott MM, Rodriguez-Mateos A, Sansone R, et al. Human red blood cells at work: identification and visualization of erythrocytic eNOS activity in health and disease. *Blood.* 2012; 120(20):4229-4237.

58. Kummer W. Pulmonary vascular innervation and its role in responses to hypoxia: size matters! *Proc Am Thorac Soc.* 2011;8(6):471-476.

59. McMahon TJ, Hood JS, Kadowitz PJ. Pulmonary vasodilator response to vagal stimulation is blocked by N omega-nitro-L-arginine methyl ester in the cat. *Circ Res.* 1992;70(2):364-369.

 Naeije R, Lejeune P, Leeman M, Melot C, Closet J. Pulmonary vascular responses to surgical chemodenervation and chemical sympathectomy in dogs. J Appl Physial (1985). 1989;66(1):42-50.
 Levitzky MG, Newell JC, Dutton RE. Effect

of chemoreceptor denervation on the pulmonary vascular response to atelectasis. *Respir Physiol.* 1978;35(1):43-51.

62. Chapleau MW, Wilson LB, Gregory TJ, Levitzky MG. Chemoreceptor stimulation interferes with regional hypoxic pulmonary vasoconstriction. *Respir Physiol.* 1988;71(2): 185-200.

63. Wilson LB, Levitzky MG. Chemoreflex blunting of hypoxic pulmonary vasoconstriction is vagally mediated. *J Appl Physiol (1985)*. 1989;66(2): 782-791.

64. Marshall JM. Peripheral chemoreceptors and cardiovascular regulation. *Physiol Rev.* 1994;74(3): 543-594.

 Lejeune P, Vachiery JL, Leeman M, et al. Absence of parasympathetic control of pulmonary vascular pressure-flow plots in hyperoxic and hypoxic dogs. *Respir Physiol.* 1989;78(2):123-133.
 Bärtsch P, Mairbäurl H, Maggiorini M, Swenson ER. Physiological aspects of high-altitude pulmonary edema. *J Appl Physiol (1985).* 2005; 98(3):1101-1110.

67. Koyama S, Kobayashi T, Kubo K, et al. The increased sympathoadrenal activity in patients with high altitude pulmonary edema is centrally mediated. *Jpn J Med.* 1988;27(1):10-16.

68. Duplain H, Vollenweider L, Delabays A, Nicod P, Bartsch P, Scherrer U. Augmented sympathetic activation during short-term hypoxia and high-altitude exposure in subjects susceptible to high-altitude pulmonary edema. *Circulation.* 1999; 99(13):1713-1718.

69. Lodato RF, Michael JR, Murray PA. Absence of neural modulation of hypoxic pulmonary vasoconstriction in conscious dogs. *J Appl Physiol (1985)*. 1988;65(4):1481-1487.

70. Albert TJ, Swenson ER. Peripheral chemoreceptor responsiveness and hypoxic pulmonary vasoconstriction in humans. *High Alt Med Biol.* 2014 (in press). Lahm T, Albrecht M, Fisher AJ, et al. 17β-Estradiol attenuates hypoxic pulmonary hypertension via estrogen receptor-mediated effects. *Am J Respir Crit Care Med.* 2012;185(9):965-980.
 Cargill RI, Lipworth BJ. Acute effects of ANP and BNP on hypoxic pulmonary vasoconstriction in humans. *Br J Clin Pharmacol.* 1995; 40(6):585-590.

73. Hubloue I, Rondelet B, Kerbaul F, et al. Endogenous angiotensin II in the regulation of hypoxic pulmonary vasoconstriction in anaesthetized dogs. *Crit Care.* 2004;8(4):R163-R171.

Thomas T, Marshall JM. The role of adenosine in hypoxic pulmonary vasoconstriction in the anaesthetized rat. *Exp Physiol.* 1993;78(4):541-543.
 Herget J, Frydrychova M, Kawikova I, McMurtry IF. Thyroxine treatment increases the hypoxic pulmonary vasoconstriction in isolated lungs from thyroidectomized rats. *Bull Eur Physiopathol Respir.* 1987;23(3):217-221.

76. Stobdan T, Karar J, Pasha MA. High altitude adaptation: genetic perspectives. *High Alt Med Biol.* 2008;9(2):140-147.

77. León-Velarde F, Mejía O. Gene expression in chronic high altitude diseases. *High Alt Med Biol.* 2008;9(2):130-139.

78. Kohler M, Kriemler S, Wilhelm EM, Brunner-LaRocca H, Zehnder M, Bloch KE. Children at high altitude have less nocturnal periodic breathing than adults. *Eur Respir J.* 2008; 32(1):189-197.

 Scoggin CH, Hyers TM, Reeves JT, Grover RF. High-altitude pulmonary edema in the children and young adults of Leadville, Colorado. *N Engl J Med.* 1977;297(23):1269-1272.
 Lorenzo VF, Yang Y, Simonson TS, et al. Genetic adaptation to extreme hypoxia: study of high-altitude pulmonary edema in a threegeneration Han Chinese family. *Blood Cells Mol Dis.* 2009;43(3):221-225.

81. Grünig E, Weissmann S, Ehlken N, et al. Stress Doppler echocardiography in relatives of patients with idiopathic and familial pulmonary arterial hypertension: results of a multicenter European analysis of pulmonary artery pressure response to exercise and hypoxia. *Circulation*. 2009; 119(13):1747-1757.

82. Balanos GM, Talbot NP, Dorrington KL, Robbins PA. Human pulmonary vascular response to 4 h of hypercapnia and hypocapnia measured using Doppler echocardiography. *J Appl Physiol* (1985). 2003;94(4):1543-1551.

83. Ketabchi F, Egemnazarov B, Schermuly RT, et al. Effects of hypercapnia with and without acidosis on hypoxic pulmonary vasoconstriction. *Am J Physiol Lung Cell Mol Physiol.* 2009;297(5):L977-L983.

 Domino KB, Pinsky MR. Effect of positive end-expiratory pressure on hypoxic pulmonary vasoconstriction in the dog. *Am J Physiol* 1990;259: 4697-705.

85. Ozaki M, Marshall C, Amaki Y, Marshall BE. Role of wall tension in hypoxic responses of isolated rat pulmonary arteries. *Am J Physiol.* 1998; 275(6 Pt 1):L1069-L1077.

86. Tsai BM, Wang M, Pitcher JM, Meldrum KK, Meldrum DR. Hypoxic pulmonary vasoconstriction and pulmonary artery tissue cytokine

Downloaded from https://prime-pdf-watermark.prime-prod.pubfactory.com/ at 2025-06-24 via free access

expression are mediated by protein kinase C. Am J Physiol. 2004;287(6):L1215-L1219.

87. Petersen B, Austen KF, Bloch KD, et al. Cysteinyl leukotrienes impair hypoxic pulmonary vasoconstriction in endotoxemic mice. *Anesthesiology*. 2011;115(4):804-811.

88. Johnson D, Hurst T, Wilson T, et al. NG-monomethyl-L-arginine does not restore loss of hypoxic pulmonary vasoconstriction induced by TNF-alpha. *J Appl Physiol (1985)*. 1993;75(2): 618-625.

89. Savale L, Tu L, Rideau D, et al. Impact of interleukin-6 on hypoxia-induced pulmonary hypertension and lung inflammation in mice. *Respir Res.* 2009;10:6.

90. Petersen B, Bloch KD, Ichinose F, et al. Activation of Toll-like receptor 2 impairs hypoxic pulmonary vasoconstriction in mice. *Am J Physiol.* 2008;294(2):L300-L308.

91. Semenza GL. Hypoxia-inducible factors in physiology and medicine. *Cell.* 2012;148(3): 399-408.

92. Engebretsen BJ, Irwin D, Valdez ME, O'Donovan MK, Tucker A, van Patot MT. Acute hypobaric hypoxia (5486 m) induces greater pulmonary HIF-1 activation in hilltop compared to madison rats. *High Alt Med Biol.* 2007;8(4): 812-821.

93. Shimoda LA, Manalo DJ, Sham JS, Semenza GL, Sylvester JT. Partial HIF-1alpha deficiency impairs pulmonary arterial myocyte electrophysiological responses to hypoxia. *Am J Physiol Lung Cell Mol Physiol.* 2001;281(1): L202-L208.

94. Yu AY, Shimoda LA, Iyer NV, et al. Impaired physiological responses to chronic hypoxia in mice partially deficient for hypoxiainducible factor 1alpha. *J Clin Invest*. 1999;103(5): 691-696.

95. Abud EM, Maylor J, Undem C, et al. 2012. Digoxin inhibits development of hypoxic pulmonary hypertension in mice. *Proc Natl Acad Sci U S A*. 2012;109(4):1239-1244.

96. Zhang H, Qian DZ, Tan YS, et al. Digoxin and other cardiac glycosides inhibit HIF-1alpha synthesis and block tumor growth. *Proc Natl Acad Sci U S A.* 2008;105(50):19579-19586.

97. Wang J, Weigand L, Lu W, Sylvester JT, Semenza GL, Shimoda LA. Hypoxia inducible factor 1 mediates hypoxia-induced TRPC expression and elevated intracellular Ca2+ in pulmonary arterial smooth muscle cells. *Circ Res.* 2006;98(12):1528-1537.

98. Smith TG, Balanos GM, Croft QP, et al. The increase in pulmonary arterial pressure caused by hypoxia depends on iron status. *J Physiol.* 2008; 586(Pt 24):5999-6005. 99. Smith TG, Talbot NP, Privat C, et al. Effects of iron supplementation and depletion on hypoxic pulmonary vasoconstriction. *JAMA*. 2009; 302(13):1444-1450.

100. Knowles HJ, Raval RR, Harris AL, Ratcliffe PJ. Effect of ascorbate on the activity of hypoxia-inducible factor in cancer cells. *Cancer Res.* 2003;63(8):1764-1768.

101. Dorrington KL, Balanos GM, Talbot NP, Robbins PA. Extent to which pulmonary vascular responses to PCO2 and PO2 play a functional role within the healthy human lung. *J Appl Physiol (1985).* 2010;108(5):1084-1096.

102. Naeije R, Brimioulle S. Physiology in medicine: importance of hypoxic pulmonary vasoconstriction in maintaining arterial oxygenation during acute respiratory failure. *Crit Care*. 2001; 5(2):67-71.

103. Gao Y, Raj JU. Regulation of the pulmonary circulation in the fetus and newborn. *Physiol Rev.* 2010;90(4):1291-1335.

104. Morin FC 3rd, Egan EA. Pulmonary hemodynamics in fetal lambs during development at normal and increased oxygen tension. *J Appl Physiol (1985).* 1992;73(1):213-218.

105. Groves BM, Droma T, Sutton JR, et al. Minimal hypoxic pulmonary hypertension in normal Tibetans at 3,658 m. *J Appl Physiol (1985)*. 1993;74(1):312-318.

106. Moore LG, Niermeyer S, Zamudio S. Human adaptation to high altitude: regional and life-cycle perspectives. *Am J Phys Anthropol.* 1998; Suppl 27:25-64.

107. Wang Z, Chesler NC. Pulmonary vascular mechanics: important contributors to the increased right ventricular afterload of pulmonary hypertension. *Exp Physiol.* 2013;98(8): 1267-1273.

108. Gomez-Arroyo J, Saleem SJ, Mizuno S, et al. A brief overview of mouse models of pulmonary arterial hypertension: problems and prospects. *Am J Physiol Lung Cell Mol Physiol.* 2012;302(10):L977-L991.

109. Barbera JA, Ramirez J, Roca J, Wagner PD, Sanchez-Lloret J, Rodriguez-Roisin R. Lung structure and gas exchange in mild chronic obstructive pulmonary disease. *Am Rev Respir Dis.* 1990;141(4 Pt 1):895-901.

110. Wagner PD, Dantzker DR, Dueck R, Clausen JL, West JB. Ventilation-perfusion inequality in chronic obstructive pulmonary disease. *J Clin Invest*. 1977;59(2):203-216.

111. Lejuene P, Mois P, Naeije R, Hallemans R, Melot C. Acute hemodynamic effects of controlled oxygen therapy in decompensated chronic obstructive pulmonary disease. *Crit Care Med.* 1984;12(12):1032-1035. 112. Timms RM, Khaja FU, Williams GW. Hemodynamic response to oxygen therapy in chronic obstructive pulmonary disease. *Ann Intern Med.* 1985;102(1):29-36.

113. Robinson TD, Freiberg DB, Regnis JA, Young IH. The role of hypoventilation and ventilation-perfusion redistribution in oxygeninduced hypercapnia during acute exacerbations of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2000;161(5):1524-1529.

114. Continuous or nocturnal oxygen therapy in hypoxemic chronic obstructive lung disease: a clinical trial. Nocturnal Oxygen Therapy Trial Group. *Ann Intern Med.* 1980;93(3):391-398.
115. Ashutosh K, Mead G, Dunsky M. Early effects of oxygen administration and prognosis in chronic obstructive pulmonary disease and cor pulmonale. *Am Rev Respir Dis.* 1983;127(4): 399-404.

116. Wagner PD, Dantzker DR, Dueck R, de Polo JL, Wasserman K, West JB. Distribution of ventilation-perfusion ratios in patients with interstitial lung disease. *Chest.* 1976;69(2 Suppl): 256-257.

 Agustí AG, Roca J, Gea J, Wagner PD, Xaubet A, Rodriguez-Roisin R. Mechanisms of gas-exchange impairment in idiopathic pulmonary fibrosis. *Am Rev Respir Dis.* 1991;143(2):219-225.
 Crockett AJ, Cranston JM, Antic N. Domiciliary oxygen for interstitial lung disease. *Cochrane Database Syst Rev.* 2001;(3):CD002883.

119. Agustí AG, Barberà JA. Contribution of multiple inert gas elimination technique to pulmonary medicine. 2. Chronic pulmonary diseases: chronic obstructive pulmonary disease and idiopathic pulmonary fibrosis. *Thorax.* 1994;49(9): 924-932.

120. Teppema LJ, Balanos GM, Steinback CD, et al. Effects of acetazolamide on ventilatory, cerebrovascular, and pulmonary vascular responses to hypoxia. *Am J Respir Crit Care Med.* 2007;175(3): 277-281.

121. Blanco I, Gimeno E, Munoz PA, et al. Hemodynamic and gas exchange effects of sildenafil in patients with chronic obstructive pulmonary disease and pulmonary hypertension. *Am J Respir Crit Care Med.* 2010;181(3):270-278.

122. Dernaika TA, Beavin M, Kinasewitz GT. Iloprost improves gas exchange and exercise tolerance in patients with pulmonary hypertension and chronic obstructive pulmonary disease. *Respiration.* 2010;79(5):377-382.

123. Boeck L, Tamm M, Grendelmeier P, Stolz D. Acute effects of aerosolized iloprost in COPD related pulmonary hypertension – a randomized controlled crossover trial. *PLoS One.* 2012;7(12): e52248.